CFD Prediction of Heterogeneities in the Scale up of Liquid-Liquid Dispersions

R. Gelves and L. Niño

Abstract—In this work simulations were developed in order to allow the examinations of heterogeneities in the scale up of liquid-liquid dispersions (oil-water) in 3 and 300 L stirred tank bioreactors using CFD simulations (Computational Fluid Dynamics). The effects of turbulence, rotating flow, drop breakage were simulated by using the k-e, MRF (Multiple Reference Frame) and PBM (Population Balance Model), respectively, providing detailed information of important bioreactor conditions. CFD modeling predicts with good approach hydrodynamic trends which can affect the liquid-liquid dispersion process of large scale bioreactors. Motivated by these results, CFD simulations are qualified as a very promising tool for predicting hydrodynamics and drop sizes especially useful for liquid-liquid applications which are characterized by the challenging problem of emulsion stability due to undesired drop and heterogeneities in the scale up process.

Index Terms—Computational fluid dynamics, drop size, population balance model, bioreactor, scale up.

I. INTRODUCTION

The liquid-liquid dispersions in turbulent flow are common in many applications, including chemical, petroleum, pharmaceutical and food industries [1]. In these bioprocesses phase dispersion and drop size have a significant importance on emulsion stability. Hence, the bioreactor operational conditions is of great importance for producing acceptable drop size to ensure stability, especially when emulsion processes are planned in large scale [2]. The loss of complete mixing conditions with increasing scale could generate gradients leading to a departure from optimal conditions found in laboratory scale. Sharp gradients of liquid-liquid dispersions are caused by poor mixing generated by empirical methods adopting as scale up strategies. All empirical scale up criteria are based on ideal approximations to maintain constant an operating parameter, leaving aside the hydrodynamics and transport phenomena governing the scale up process. Knowing the hydrodynamic behavior on the scale up bioreactors allows identifying the degree of departure from perfect mixing conditions associated with scaling rules.

Drop breakage and coalescence during high-shear preparation have a substantial impact on the drop size distribution. While drop breakage conditions have been extensively studied, predictive models for hydrodynamics, drop breakage and coalescence under turbulent conditions are lacking in a scale up process [3]. Due the lack of suitable

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models, emulsified products in the scale up liquid-liquid dispersions are currently developed by combining a broad knowledge of the previous product formulations with empirical scientific experimentation. Because this approach is intuitive and experimental, the progression of a formulation is generally unpredictable and a new product will often go through hundreds of prototype formulations in a laboratory or pilot plant before commercialization [3]. Due to the very large number of possible formulation and processing combinations that need to be explored in a scale up process, the traditional trial-and-error approach requires significant time and resources [3]. Hence it is the motivation of this work to present a CFD simulation approach for the estimation of drop size and heterogeneities due to the scale up process in liquid-liquid systems.

II. METHODS

A 3 L stirrer tank with 0.160 m diameter equipped with a 0,059 m Rushton turbine diameter and two cylindrical baffles were used for the experiments at lab scale and different stirrer speeds (350, 500 and 650 rpm) were tested for defining the best condition [2] to implement in further scale up. It was used sunflower oil as dispersed phase. It was mainly used at concentration of 10% (v/v) in de-ionized water at atmospheric pressure and a constant temperature of 20 °C. Coalescence was reduced by sodium dodecyl sulfate - SDS provided by Sygma-Aldrich Co. An endoscope technique was developed [4], [5]. Considering this technique, drop size distributions for all phase fractions even under transient conditions [6], [7] can be determined with high time resolution.

The drops are semi-automatically measured and counted. The technique is capable of measuring drops of 25 μ m to 1000 μ m at any dispersed phase fraction. This technique offers reliable in-situ measurement of drop sizes from any part of the stirred vessel [5]. The endoscope technique (developed by Maass [5]) was applied in order to measure the drop size distributions as a function of parameters like stirrer speed.

Besides the general technical importance of analyzing the influence of these parameters, this set-up is very instructive for the analysis of the physical phenomena in stirred dispersions in a technically relevant parameter range. This is a major requirement for the development of CFD models for immiscible liquids dispersions [5].

For simulating heterogeneities in the scale up stage, a 300 L virtual bioreactor was dimensioned (0.57 m diameter) maintaining a geometric similarity from the tested 3 L bioreactor. Operational conditions were defined based on the best condition found in lab scale bioreactor (3 L) and using

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the P/V (constant power input per liquid volume) empirical scale up criterion [8].

The Eulerian model is the most complex multiphase model in ANSYS FLUENT 13.0. It solves a system of *n*-momentum and continuity equations for each phase. The coupling is achieved through pressure and interfacial exchange coefficients. The mass conservation equation for each phase is shown below:

$$\frac{\partial}{\partial t}(\rho_i \alpha_i) + \nabla \cdot \left(\alpha_i \rho_i \vec{U}_i\right) \tag{1}$$

where ρ_i , α_i and \vec{U}_i represent the density, volume fraction and the rate of phase *i* (oil or water). It is assumed that the water phase and the oil phase share space in proportion to their volume, such that their volume fractions sums to unity in the cell domain.

$$\alpha_G + \alpha_L = 1.0 \tag{2}$$

The momentum equation for phase *i* is described below:

$$\frac{\partial}{\partial t} (\rho_i \alpha_i \vec{U}_i) + \nabla \cdot (\alpha_i \rho_i \vec{U}_i \vec{U}_i) = \alpha_i \nabla p
+ \nabla \cdot \overline{\tau}_{effi} + \vec{R}_i + \vec{F}_i + \alpha_i \rho_i \vec{g}$$
(3)

p is the pressure shared by both phases and $\vec{R_i}$ represents the interfacial momentum exchange. The $\vec{F_i}$ term represents the Coriolis and centrifugal forces expressed in the MRF model for rotating flows and is represented as:

$$\vec{F}_i = -2\alpha_i \rho_i \vec{N} \times \vec{U}_i - \alpha_i \rho_i \vec{N} \times \left(\vec{N} \times \vec{r}\right)$$
(4)

 \vec{N} is the angular velocity, \vec{r} is the position vector. The Reynolds stress tensor $\bar{\bar{\tau}}_{effi}$ is related to the mean velocity gradients through the Boussinesq hypothesis [9]:

$$\bar{\tau}_{effi} = \alpha_i (\mu_{lam,i} + \mu_{t,i}) (\nabla \vec{U}_i + \nabla \vec{U}_i^T) - \frac{2}{3} \alpha_i (\rho_i k_i + (\mu_{lam,i} + \mu_{t,i}) \nabla \cdot \vec{U}_i) \bar{I}$$
(5)

 $\mu_{lam,i}$ is the molecular viscosity of phase *i*, $\overline{\overline{I}}$, is the strain tensor.

The most important force exchange between the phases is drag force acting on drops. This force depends on friction, pressure, cohesion, and other hydrodynamic effects [10].

$$R_L = -R_G = K \left(\vec{U}_G - \vec{U}_L \right) \tag{6}$$

K is the exchange coefficient of water and oil phases and is determined by the equation:

$$K = \frac{3}{4}\rho_L \alpha_L \alpha_G \frac{C_D}{d} \left| \vec{U}_G - \vec{U}_L \right| \tag{7}$$

d is the drop diameter and the drag coefficient and C_D is defined as a function of Reynolds number:

$$Re_p = \frac{\rho_L |\vec{U}_G - \vec{U}_L| d}{\mu_L} \tag{8}$$

To calculate the drag coefficient using the standard correlation [11]:

$$C_D = \begin{cases} \frac{24(1+0.15Re_p^{0.687})}{Re_p}, & Re_p \le 1000\\ 0.44, & Re_p > 1000 \end{cases}$$
(9)

The dispersed turbulence $k - \varepsilon$ model can be considered as the multiphase standard turbulence model. It represents the extension of the single phase $k - \varepsilon$ model and is used when the secondary phase concentrations are diluted on primary phase. k and ε equations describing this model are as follows:

$$\frac{\partial}{\partial t} (\rho_L \alpha_L k_L) + \nabla \cdot (\rho_L \alpha_L \vec{U}_L k_L)
= \nabla \cdot \left(\alpha_L \frac{\mu_{t,L} \nabla k_L}{\sigma k} \right) + \alpha_L G_{k,L} - \alpha_L \rho_L \varepsilon_L
+ \alpha_L \rho_L \Pi_{K,L}$$
(10)

$$\frac{\partial}{\partial t} (\rho_L \alpha_L k_L \varepsilon_L) + \nabla \cdot (\rho_L \alpha_L \vec{U}_L \varepsilon_L)
= \nabla \cdot \left(\alpha_L \frac{\mu_{t,L} \nabla \varepsilon_L}{\sigma \varepsilon} \right) + \alpha_L \frac{\varepsilon_L}{k_L} (C_{1\varepsilon} G_{k,L} - C_{2\varepsilon} \rho_L \varepsilon_L)
+ \alpha_L \rho_L \Pi_{\varepsilon,L}$$
(11)

In these equations, $G_{k,L}$ represents the generation of turbulent kinetic energy k_L of the water phase due to mean velocity gradients, ε_L is the turbulent dissipation energy. $\Pi_{K,L}$ and $\Pi_{\varepsilon,L}$ represent the influence of the dispersed oil phase in the continuous phase and are modeled with the Elgobashi and Rizk [7] equations.

The turbulent viscosity $\mu_{t,L}$, is calculated from:

$$\mu_{t,L} = \rho_L C_\mu \frac{k_L^2}{\varepsilon_L} \tag{12}$$

The values of the constants used in this experiment were $C_{1\varepsilon}$: 1.44, $C_{2\varepsilon}$: 1.92, C_{μ} :0.09 σ_k : 1.00 and σ_{ε} : 1.30. σ_k and σ_{ε} represent turbulent Prandtl number for k and ε , respectively.

The eulerian approach was coupled to population balance models. The finite volume technique was used in Ansys Fluent 13 to solve the equations. The effects of turbulence and rotating flow were simulated by using the k-e and MRF (Multiple Reference Frame) models. The discrete method [12] is used in this manuscript to solve the population balance equations. The drop population is discretized into a finite number of intervals of drop sizes. The population balance equations for different drop classes can be written as [13]:

$$\frac{\partial}{\partial t}(\rho_G n_i) + \nabla \cdot \left(\rho_G \vec{U}_G n_i\right) = \rho_G \left(\Gamma_{B_{i_B}} - \Gamma_{D_{i_B}}\right) \quad (13)$$

where n_i is the number of classes of drop *i*, $\Gamma_{B_{i_B}}$ is the birth rate due to breakage and $\Gamma_{D_{i_B}}$ is the death rate. The terms of breakage are:

$$\Gamma_{B_{i_B}} = \int_{\Omega_{v}} pg(v')\beta(v|v')n(v',t)dv'$$
(14)

$$\Gamma_{D_{i_R}} = g(v)n(v,t) \tag{15}$$

g(v) is the breakup rate of drops of size v; g(v'), is the breakup frequency of drop v' and $\beta(v|v')$ is the probability density function of drops broken from the volume v' in a drop of volume v. The drop breakup is analyzed in terms of drops interaction with turbulent eddies. These turbulent eddies increase the drop surface energy to cause deformation. The breakup occurs if the increase in the surface energy reaches a critical value. The breakup rate is defined as [14]:

$$g(v')\beta(v|v') = k \int_{\xi_{min}}^{1} \frac{(1+\xi)^2}{\xi^{11/3}} exp\left(-b\xi^{-11/3}\right) d\xi \quad (16)$$

$$k = 0.9238\varepsilon^{1/3} d^{-2/3} \alpha \tag{17}$$

$$b = 12 \left(f^{2/3+} (1-f)^{2/3} - 1 \right) \sigma \rho^{-1} \varepsilon^{-2/3} d^{-5/3}$$
(18)

where d is the particle diameter, ξ is the dimensionless eddy size, f is the breakage frequency.

The 3D mesh is composed for hybrid cells with 25000 and 450000 computational cells for 3 and 300 L bioreactors, respectively (Fig. 1 and Fig. 2). The finite volume technique implemented in the CFD code Ansys Fluent 13.0 Software was used to convert the Navier- Stokes equations into algebraic equations which can be solved numerically. Tank walls, stirrer surfaces and baffles are treated with no slip conditions and standard wall functions.



Fig. 1. Mesh generation for the liquid-liquid system (3 L bioreactor).

To solve the partial differential equations the PC SIMPLE algorithm was used which couples pressure and velocity. The second order Upwind scheme was applied for the spatial terms. It was assumed that the solution converges when the scaled residuals remain with values smaller than 10⁻⁵ and when the pseudo-regime for sauter diameter is reached.



Fig. 2. Mesh generation for the liquid-liquid system (300 L bioreactor).

III. RESULTS

The mean goal of this work was to study the heterogeneities due to loss of complete mixing conditions with increasing scale, using CFD simulations. Special emphasis was given to the elucidation of liquid-liquid volume fraction dispersion and drop size distributions. Operational conditions were defined based on the best condition found in the 3L lab scale bioreactor [2] and using the P/V (constant power input per liquid volume) empirical scale up criterion. Fig. 3 and Fig. 4 show the oil volume fractions for the liquid-liquid system in 3 and 300 L bioreactor simulated by CFD.



Fig. 3. Oil volume fractions [-] for the liquid-liquid system (3 L bioreactor).



Fig. 4. Oil volume fractions [-] for the liquid-liquid system (300 L bioreactor).

Simulating the scale up in 300 L, it can be seen that relatively low oil dispersion occurs especially in bottom of bioreactor because of low centrifugal forces generated by the Rushton turbine. Obviously, centrifugal forces and turbulence dissipation energies could not overcome the oil immiscibility in water and the appearance of heterogeneous environment is becoming important for the liquid-liquid dispersion because emulsion stability can be affected. Contrarily, oil gradients are minimal when the stirrer tank is operated at lab scale (3 L) due to increase in turbulence dissipation energy [2].

Moreover, Fig. 5 and Fig. 6 depict another reason for explaining the high discrepancy between 3 and 300 L oil vol ume fraction dispersion. Turbulence dissipation energy profiles in both scales are significantly different.





Fig. 7 and Fig. 8 show the velocity magnitude calculated





Fig. 5. Turbulence dissipation energy [m²/s³] for the liquid-liquid system (3 L bioreactor) [17].



Fig. 6. Turbulence dissipation energy $[m^2/s^3]$ for the liquid-liquid system (300 L bioreactor).

While the lab scale stirred tank bioreactor shows well-defined turbulence dissipation energy zones between the impeller discharge region, walls and baffles, the 300 L bioreactor shows dead zones on bottom bioreactor. Consequently, poor mixed lower region of the bioreactor are more significant due to loss of complete mixing conditions with increasing scale.

Fig. 7. Oil velocity magnitude [m/s] for the liquid-liquid system (3 L bioreactor) [17].



Fig. 8. Oil velocity magnitude [m/s] for the liquid-liquid system (300 L bioreactor).

The different liquid velocity contours are also checked by the analysis of Kolmogorov length scale λ_K [15]:

$$\lambda_K = \left(\frac{\varepsilon_L}{v^3}\right)^{-\frac{1}{4}} \tag{19}$$

with ε_L being the turbulence dissipation energy calculated from CFD and v the dynamic viscosity of the liquid. Following this definition, the Kolmogorov diameter investigates eddies, created at the "hot spots" of energy input [10]. According to the hypothesis, only eddies with length scale smaller than drop diameters can induce breakage. Larger eddies lead to drop transport instead [14]. Fig. 9 and Fig. 10 show the Kolmogorov length scale λ_K simulated by CFD in both scales (3 and 300L). Considering these results, smaller length scales, which dominate high breakage phenomenon, are present in lab scale bioreactor than 300 L bioreactor. This finding can also explain the heterogeneities found at 300 L scale.





Fig. 9. Kolmogorov length scale [m] for the liquid-liquid system (3 L bioreactor).



Fig. 10. Kolmogorov length scale [m] for the liquid-liquid system (300 L bioreactor).

In these bioprocesses phase dispersion and drop size have a significant importance on emulsion stability. Hence, the bioreactor operational conditions are of great importance for producing acceptable drop size to ensure stability, especially when emulsion processes are planned in large scale. The most relevant parameter for analyzing these hydrodynamic mechanisms is the sauter diameter. Fig. 11 and Fig. 12 show the sauter mean diameter distribution simulated using CFD in both scales (3 and 300L).





Fig. 11. Sauter mean diameter [m] for the liquid-liquid system (3 L bioreactor).



Fig. 12. Sauter mean diameter [m] for the liquid-liquid system (300 L bioreactor).

It can be interpreted as a parameter, summarizing all individual impacts affecting the drop sizes and their distributions in one single value [10]. It is estimated coupling CFD-based hydrodynamics with population balance models (see equation (13)). Interestingly, lowest values of ~136 μ m were found close to the blades in the cross-sectional areas of the agitator speed direction. These regions mirror relatively high, local drop breakage rates.

In the case of 300 L bioreactor (Fig. 12) the sauter diameter

was higher (229-235 μ m) than the values found for 3 L bioreactor (Fig. 11). Besides, the 300L bioreactor reveals a significantly non-homogeneous sauter mean diameter distribution, while special differences are minimal using the 3 L bioreactor. The latter is the consequence of the improved hydrodynamics which promotes the formation of uniform bubble sizes, generally caused by the mixing.

Moreover, experimental sauter diameter was performed at 3 L scales [2] using an endoscope prove and compared to CFD results (Table I).

TABLE I: CFD AND EXPERIMENTAL RESULTS (3 L)

rpm	350		500		650	
d ₃₂	CFD	Exp	CFD	Exp	CFD	Exp
[µm]	248	242	145	151	136	122

Interestingly, CFD simulations show that lowest drop size values are reached at 650 rpm (137 μ m). Independent drop size measurements by an experimental endoscope technique applying the same operational conditions confirmed these findings [2]. For that reason 650 rpm was chosen in this work at starting point for calculating the operational conditions at 300 L based on P/V constant. The very good agreement is qualified as an evidence for suitability of the breakage terms used in CFD to formulate the population balance models. Besides it provides the sound basis for the discussion of the oil volume contour plots and the resulting d_{32} distributions. Both represent the backbone for the comparison of the emulsion stability not only for lab but also for large scale bioreactors.

IV. CONCLUSION

The numerical results from a scale up process of liquid-liquid dispersions are analyzed using CFD. Possible heterogeneities were simulated at 300 L bioreactor due to poor mixing conditions reached at this scale. Motivated by these simulated and experimental results CFD simulations are qualified as a very promising tool for predicting hydrodynamics and drop sizes especially useful for liquid-liquid applications which are characterized by the challenging problem of emulsion stability due to undesired drop sizes in large scale bioprocess.

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