

# Evaluation of Antioxidant Activity of Pomegranate Molasses by 2,2-Diphenyl-1-Picrylhydrazyl (DPPH) Method

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**Abstract**—In Turkish cuisine the pomegranate molasses (PM) are used as a condiment and believed to have significant effects for arteriosclerosis, cholesterol levels and cancer prevention due to the antioxidant potential of pomegranate fruit itself. In this study, we measured the total polyphenols content, of which varied from 118.28 to 828.15 mg of gallic acid equivalent per gram of PM, and antioxidant activity by DPPH assay, of which were found to be between 560.23 to 1885.23  $\mu\text{mol}$  trolox equivalent per gram of sample. The chemical composition of PM samples were found as: the water soluble dry matter content 62.40-75.00 g  $100\text{g}^{-1}$ ; viscosity 176 and 2900 mPa.s.; total acidity 4.70-9.73 g  $100\text{g}^{-1}$ ; pH of the samples changed 1.71 and 2.96; invert sugar and total sugar 23.71-56.95 g  $100\text{g}^{-1}$  and 30.33-70.94 g  $100\text{g}^{-1}$ , respectively.

**Index Terms**—Pomegranate molasses, antioxidant activity.

## I. INTRODUCTION

In recent years, scientific interest has focused on understanding the mechanisms of traditional remedy applications based on nutraceuticals and functional food nutrients. The pomegranate fruit (*Punica granatum* L.), one of the oldest edible fruits that widely grows in many tropical and subtropical countries has been considering as a health-promoting food [1]. It has been featured prominently in all the major religions of the world and has been used for centuries for the treatment of various ailments [2]. The unique chemistry of pomegranate tree have led many studies to be conducted to evaluate the functional efficacy of this fruit due to the claims related to wellbeing. Since the traditional importance of pomegranate as a medicinal plant is well-established by scientific researches, market demand and production of pomegranate products thereof showed a considerable increase [3]-[6]. The compounds in different parts of the fruit have been shown to possess preventive and attenuating activities against numerous chronic and health threatening maladies such as prostate cancer, atherosclerosis and cardiovascular diseases, oxidation of low density lipoprotein (LDL), high density lipoprotein (HDL) and cholesterol, type 2 diabetes, hypertension, arthritis, anemia and Alzheimer's disease [7]-[16].

The chemical composition of the pomegranate and its products depends on the cultivar, growing region and

climate, the fruit's stage of maturity, cultural practices and manufacturing systems [7], [17]-[19], [20].

The edible part of the pomegranate fruit consists of 40% arils and 10% seeds. The juice of arils (pomegranate juice) is comprised of 85% water, 10% total sugars (glucose, sucrose, and fructose), and 1.5% pectin, organic acids (citric, malic, tartaric, succinic, fumaric, ascorbic acid), fatty acids (i.e. conjugated linoleic acid, linoleic acid, punicic acid and eleostearic acid) and amino acids (i.e. proline, valin, and methionine) and bioactive compounds (phenolics and flavonoids). The seeds are a rich source of total lipids; and pomegranate seed oil comprises 12% to 20% of total seed weight. The predominant fatty acid in seed oil is 9,11,13-octadeca-trienoic acid (punicic acid), a conjugated linolenic acid characteristic from pomegranate seeds, with contents between 3523 and 10586 mg  $100\text{g}^{-1}$  of seeds [21]-[27].

Mounting evidence suggests that the beneficial health effects of fruits and vegetables in the prevention of disease are due to the bioactive compounds such as phenolic acids, flavonoids, tannins and antioxidant properties. Antioxidants are compounds that have the ability to scavenge free radicals and protect the human body from oxidative stress, the main key factor for the occurrence of cancers and heart diseases [28]-[30]. Reference [31] stated that the antioxidant level in pomegranate juice was higher than found in many fruit juices, such as blueberry, cranberry, and orange. Reference [32] and Reference [33] demonstrated that pomegranate juice and seed extracts have 2-3 fold more *in vitro* antioxidant capacity than red wine or green tea. Ellagitannins, mainly punicalagins, are reported to be responsible for over 50% of the antioxidant activity and the above health promoting features of pomegranate [34], [35].

Pomegranate can be consumed as fresh, fruit juice, fermented fruit juice, dried aril, frozen aril, minimally-processed aril, canned aril, jam, jelly, wine, vinegar, paste, fruit leather and in flavoring products. The '*pomegranate molasse*', also named as sour pomegranate pekmez, nar eksisi, pomegranate sauce, is a traditional condiment commonly used in salads and many dishes to improve the taste and aroma characteristics in Turkey. It is a concentrated product produced simply by boiling of pomegranate juice without the further addition of sugar or other additives. Traditional methods are still being used to produce pomegranate molasses, a thick, dark red liquid formed after cleaning, crushing, extraction, filtration, and evaporation in an open vessel or under vacuum (Fig. 1). Clarification step is generally omitted since customers prefer bitterness and sourness caused by phenolic substances and acidity [36]-[38].

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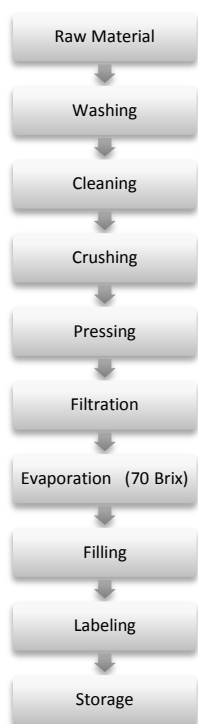


Fig. 1. The process of pomegranate molasses production.

The antioxidant activity of pomegranate fruit juice, peels, leaves, arils or seeds have been reported separately by several researchers due to their wellknown ethnomedical relevance and chemical features [33], [39]-[44], [25], [4], however, there are few reports on the chemical composition and antioxidant activity of pomegranate molasses [45], [46]. Therefore, the main objective of the present work was to determine the antioxidant activity of pomegranate molasses by the DPPH assay, as well as the total phenolics contents.

## II. MATERIALS AND METHODS

### A. Materials and Methods

Pomegranate molasses samples were purchased from a local markets in Bursa. All chemicals were analytical-reagent grade and obtained from the following sources: 2,2-diphenyl-1-picrylhydrazyl (DPPH), Folin-Ciocalteu reagent,  $\text{Na}_2\text{CO}_3$  (Sigma-Aldrich S.p.A., Milan, Italy); gallic acid and ethanol (Merck, Darmstadt, Germany).

### B. Chemical Analysis of PM

The water soluble dry matter content (brix) of the samples was expressed as  $\text{g } 100\text{g}^{-1}$  by using an Abbe refractometer [47] Viscosity was measured in mPa.s using a rotary viscosimeter (NDJ-1, Shanghai Precision & Scientific Inst. Co. Ltd.). Total acidity was determined as citric acid, by titrating samples against NaOH solution of known normality [48]. pH was measured potentiometrically by a Nel-pH 890 model pH meter. Invert and total sugars were determined by the Luff-Schoorl method [49].

### C. Analysis of Total Phenolic Content of PM

Samples were diluted to 14 brix and 1 milliliter of sample and 10 mL of extraction solution (Methanol for TPC, ethanol for AC) were mixed and shaken at  $20^\circ\text{C}$  for 10 minutes. Then the suspension was centrifuged at 3 500 rpm for 10 minutes and filtered.

The total phenolic content of pomegranate molasses was determined spectrophotometrically at 725 nm according to modified Folin–Ciocalteu colorimetric method [50]. After adding 0.1 mL extract, 2.3 mL distilled water, 0.15 mL Folin-Ciocalteu reagent and vortexed for 15 seconds. After 5 minutes 0.30 mL 35%  $\text{Na}_2\text{CO}_3$  added and content was mixed and left to stand at room temperature in dark for 120 min. Samples were measured 725 nm. A standard calibration curve was plotted using gallic acid. The results were expressed as “milligrams of gallic acid equivalents per gram of pomegranate molasses”.

### D. Measurement of Antioxidant Activity by DPPH

The antioxidant activity of pomegranate molasses were determined according to the method reported by Reference [51]. 0.1 mL prepared supernatant and 3.9 mL DPPH ( $6 \times 10^{-5}$  M) solution were mixed. The mixture was shaken vigorously and was allowed to stand at room temperature for 30 min before the decrease in absorbance at 515 nm was measured. Antioxidant activity was determined as the percentage of DPPH decrease using the equation:

$$\text{Inhibition \%} = (\text{Abs}_{t=0} - \text{Abs}_{t=30 \text{ min}}) / \text{Abs}_{t=0} \times 100$$

$\text{Abs}_{t=0}$  was the absorbance of DPPH at time 0.

$\text{Abs}_{t=30 \text{ min}}$  was the absorbance of DPPH after 30 min of incubation.

Inhibition (%) was calculated according to trolox calibration curve as “ $\mu\text{mol}$  trolox equivalent per gram of sample”.

## III. RESULTS AND DISCUSSION

Total phenolic content and antioxidant activity of pomegranate molasses were shown in Table I.

TABLE I: FUNCTIONAL PARAMETERS OF PM

Sample	Antioxidant Capacity	Total Phenolic Component
PMI	190,60 $\pm$ 0,48	344,30 $\pm$ 1,84
PMII	271,89 $\pm$ 10,52	691,25 $\pm$ 4,86
PMIII	471,85 $\pm$ 2,33	828,15 $\pm$ 6,79
PMIV	140,22 $\pm$ 2,84	168,89 $\pm$ 2,91
PMV	410,15 $\pm$ 4,60	745,46 $\pm$ 4,45
PMVI	155,34 $\pm$ 0,58	325,51 $\pm$ 0,64
PMVII	208,01 $\pm$ 3,00	118,28 $\pm$ 0,00
PMVIII	411,92 $\pm$ 3,07	711,94 $\pm$ 0,91
PMIX	222,57 $\pm$ 0,75	631,69 $\pm$ 4,08

The chemical composition of PM samples were found as: the water soluble dry matter content  $62.40\text{--}75.00 \text{ g } 100\text{g}^{-1}$ ; viscosity 176 and 2900 mPa.s.; total acidity  $4.70\text{--}9.73 \text{ g } 100\text{g}^{-1}$ ; pH of the samples changed 1.71 and 2.96; invert sugar and total sugar  $23.71\text{--}56.95 \text{ g } 100\text{g}^{-1}$  and  $30.33\text{--}70.94 \text{ g } 100\text{g}^{-1}$ , respectively (data not shown).

The total polyphenol content was calculated via the gallic acid curve of the spectrophotometric measurement result of the absorbance of the blue color at 765 nm, which is the result of the redox reaction of the reduction of the Folin-Ciocalteu reagent by phenolic compounds and turning into oxidized forms in an alkali medium. Phenolic compounds give the color, astringency and bitter taste to pomegranate. Since pomegranate molasses is a concentrated product, it was expected to have high phenolic content. However, due to thermal processing conditions (simply boiling) and

inclusion of saccharose syrup to give a caramelized flavour with artificial pomegranate flavour phenolic contents were low.

Reactive oxygen species (ROS) such as superoxide anion ( $O_2^{\bullet-}$ ), hydroxyl ( $\bullet OH$ ), peroxy ( $ROO\bullet$ ), and alkoxy radicals ( $RO\bullet$ ), hydrogen peroxide ( $H_2O_2$ ), and singlet oxygen ( $O_2^1\Delta g$ ) may attack biological macromolecules, giving rise to protein, lipid, and DNA damage, cell aging, oxidative stress-originated diseases (e.g., cardiovascular and neurodegenerative diseases), and cancer. Antioxidants, either exogenous or endogenous, are vital substances which possess the ability to scavenge or quench ROS and reactive nitrogen species (RNS) in order to protect the body from the potent injuries caused by these radicals. Measuring the antioxidant capacity/activity of any foodstuff is carried out for the meaningful comparison of the antioxidant content for the diagnosis and treatment of oxidative stress-associated diseases. The DPPH radical has widespread use in the free radical scavenging activity assessment because of the ease and convenience of the reaction.

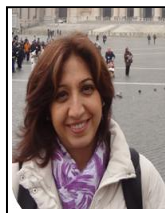
In the presence of arbutus extracts and standard solutions the absorbance of DPPH solution decreased, of which were found to be between 16.11 to 75.22%, indicating that radical scavenging activity was concentration dependent.

It could be said that the antioxidant activity of pomegranate molasses depend on several factors, such as cultivar and climatic conditions during fruit maturation and ripening and the part of the fruit used.

#### REFERENCES

- [1] M. Rinaldi, A. Caligiani, R. Borgese, G. Palla, D. Barbanti, and R. Massini, "The effect of fruit processing and enzymatic treatments on pomegranate juice composition, antioxidant activity and polyphenols content," *LWT – Food Science and Technology*, vol. 53, pp. 355–359, 2013.
- [2] D. N. Syed, F. Afaq, and H. Mukhtar, "Pomegranate derived products for cancer chemoprevention Seminars in Cancer Biology," *Seminars in Cancer Biology*, vol. 17, issue 5, pp. 377–85, 2007.
- [3] P. Langley, "Why a pomegranate?" *British Medical Journal*, vol. 321, no. 7269, pp. 1153–1154, 2000.
- [4] Z. Pan, W. Qu, H. Mab, G. G. Atungulu, and T. H. McHugh, "Continuous and pulsed ultrasound-assisted extractions of antioxidants from pomegranate peel," *Ultrasonics Sonochemistry*, vol. 18, pp. 1249–1257, 2010.
- [5] M. A. E. Ahmed, E. M. ElMorsy, and A. A. E. Ahmed, "Pomegranate extract protects against cerebral ischemia/reperfusion injury and preserves brain DNA integrity in rats," *Life Sciences*, vol. 110, pp. 61–69, 2014.
- [6] O. Orgil, E. Schwartz, L. Baruch, I. Matityahu, J. Mahajna, and R. Amir, "The antioxidative and anti-proliferative potential of non-edible organs of the pomegranate fruit and tree," *LWT – Food Science and Technology*, vol. 58, pp. 571–577, 2014.
- [7] A. Basu and K. Penugonda, "Pomegranate Juice: A heart-healthy fruit juice," *Nutrition Reviews*, vol. 67, no. 1, pp. 49–56, 2009.
- [8] A. Esmailzadeh, F. Tahbaz, I. Gaieni, H. Alavi-Majd, and L. Azadbakht, "Concentrated pomegranate juice improves lipid profiles in diabetic patients with hyperlipidemia," *J. Medic Food*, vol. 7, pp. 305–308, 2004.
- [9] D. Heber, "Multitargeted therapy of cancer by Ellagitannins," *Cancer Letters*, vol. 269, no. 2, pp. 262–268, 2008.
- [10] N. D. Kim, R. Mehta, W. Yu *et al.*, "Chemopreventive and adjuvant therapeutic potential of pomegranate (*Punica granatum*) for human breast cancer," *Breast Can Res Treat*, vol. 71, pp. 203–217, 2002.
- [11] M. Rosenblat, N. Volkova, R. Coleman, and M. Aviram, "Pomegranate by-product administration to apolipoprotein e-deficient mice attenuates atherosclerosis development as a result of decreased macrophage oxidative stress and reduced cellular uptake of oxidized low-density lipoprotein," *J. Agric Food Chem*, vol. 54, pp. 1928–1935, 2006.
- [12] E. P. Lansky and R. A. Newman, "Punica granatum (pomegranate) and its potential for prevention and treatment of inflammation and cancer," *J. Ethnopharmacol*, vol. 109, pp. 177–206, 2007.
- [13] M. R. Sartippour, N. P. Seeram, J. Y. Rao *et al.*, "Ellagitannin-rich pomegranate extract inhibits angiogenesis in prostate cancer in vitro and in vivo," *International Journal of Oncology*, vol. 32, no. 2, pp. 475–480, 2008.
- [14] M. Viuda-Martos, M. C. Lopez-Marcos, J. Fernandez-Lopez *et al.*, "The role of fiber in cardiovascular diseases: A review," *Comp Rev Food Sci Food Safety*, vol. 9, pp. 240–258, 2010.
- [15] K. Al-Jarallah, D. Shehab, W. Al-Azmi, and A. Al-Fadli, "Rheumatic complications of inflammatory bowel disease among Arabs: a hospital-based study in Kuwait," *International Journal of Rheumatic Diseases*, vol. 16, no. 2, pp. 134–138, 2013.
- [16] S. Banihani, S. Swedan, and Z. Alguraan, "Pomegranate and type 2 diabetes," *Nutr Res*, vol. 33, pp. 341–348, 2013.
- [17] R. K. Toor, G. P. Savage, and C. E. Lister, "Seasonal variations in the antioxidant composition of greenhouse-grown tomatoes," *Journal of Food Composition and Analysis*, vol. 19, no. 1, pp. 1–10, 2006.
- [18] A. Raffo, G. La Malfa, V. Fogliano, G. Madani, and G. Quaglia, "Seasonal variations in antioxidant components of cherry tomatoes (*Lycopersicon esculentum* cv. Naomi)," *Journal of Food Composition and Analysis*, vol. 19, no. 1, pp. 11–19, 2006.
- [19] H. Borochoy-Neori, S. Judeinstein, E. Tripler, M. Harari, A. Greenberg, I. Shomer, and D. Holland, "Seasonal and cultivar variations in antioxidant and sensory quality of pomegranate (*Punica granatum* L.) fruit," *Journal of Food Composition and Analysis*, vol. 22, no. 3, pp. 189–195, 2009.
- [20] M. Zarei, M. Azizi, and Z. Bashir-Sadr, "Evaluation of physicochemical characteristics of pomegranate (*Punica granatum* L.) fruit during ripening," *Fruits*, vol. 66, no. 2, pp. 121–129, 2011.
- [21] M. Aviram, L. Dornfeld, M. Rosenblat *et al.*, "Pomegranate juice consumption reduces oxidative stress, atherogenic modifications to LDL, and platelet aggregation: Studies in humans and in atherosclerotic apolipoprotein E-deficient mice," *Am J Clin Nutr*, vol. 71, pp. 1062–1076, 2000.
- [22] Y. Li, C. Guo, J. Yang, J. Wei, J. Xu, and S. Cheng, "Evaluation of antioxidant properties of pomegranate peel extract in comparison with pomegranate pulp extract," *Food Chem*, vol. 96, no. 2, pp. 254–260, 2006.
- [23] A. Fadavi, M. Barzegar, and H. M. Azizi, "Determination of fatty acids and total lipid content in oilseed of 25 pomegranates varieties grown in Iran," *J Food Comp Anal*, vol. 19, pp. 676–680, 2006.
- [24] S. H. Mirdehghan and M. Rahemi, "Seasonal changes of mineral nutrients and phenolics in pomegranate (*Punica granatum* L.) fruit," *Sci Hort*, vol. 111, no. 2, pp. 120–127, 2007.
- [25] F. Tezcan, M. Gultekin-Özügen, T. Diken, B. Özgelik, and F. B. Erim, "Antioxidant activity and total phenolic, organic acid and sugar content in commercial pomegranate juices," *Food Chem*, vol. 115, no. 3, pp. 873–877, 2009.
- [26] S. S. Dahham, M. N. Ali, H. Tabassum, and M. Khan, "Studies on Antibacterial and Antifungal Activity of Pomegranate (*Punica granatum* L.)," *American-Eurasian J. Agric. & Environ. Sci.*, vol. 9, no. 3, pp. 273–281, 2010.
- [27] L. Fernandes, J. A. Pereira, I. Lopez-Cortes, D. M. Salazar, E. Ramalhosa, and S. Casal, "Fatty acid, vitamin E and sterols composition of seed oils from nine different pomegranate (*Punica granatum* L.) cultivars grown in Spain," *Journal of Food Comp and Analysis*, vol. 39, pp. 13–22, 2005.
- [28] R. Amarowicz, R. B. Pegg, R. Rahimi-Moghaddam, B. Barl, and J. A. Weil, "Free-radical scavenging capacity and antioxidant activity of selected plant species from the Canadian prairies," *Food Chem*, vol. 84, pp. 551–562, 2004.
- [29] A. Scalbert, C. Manach, C. Morand, C. Remesy, and L. Jimenez, "Dietary polyphenols and the prevention of diseases," *Crit Rev Food Sci Nut*, vol. 45, pp. 287–306, 2005.
- [30] R. Tzulkar, I. Glazer, I. Bar-Ilan, D. Holland, M. Aviram, and R. Amir, "Antioxidant activity, polyphenol content and related compounds in different fruit juices and homogenates prepared from 29 different pomegranate accessions," *J Agric Food Chem*, vol. 55, pp. 9559–9570, 2007.
- [31] N. P. Seeram, L. S. Adams, S. M. Henning *et al.*, "In vitro antiproliferative, apoptotic and antioxidant activities of punicalagin, ellagic acid and a total pomegranate tannin extract are enhanced in combination with other polyphenols as found in pomegranate juice," *Journal of Nutritional Biochemistry*, vol. 16, no. 6, pp. 360–367, 2005.
- [32] S. Y. Schubert, E. P. Lansky, and I. Necman, "Antioxidant and eicosanoid enzyme inhibition properties of pomegranate seed oil and fermented juice flavonoids," *Journal of Ethnopharmacology*, vol. 66, no. 1, pp. 11–17, 1999.

- [33] M. Gil, I. Tomas-Barberan, F. Hess-Pierce, B. D. M. Holcroft, and A. Kader, "Antioxidant activity of pomegranate juice and its relationship with phenolic composition and processing," *Journal of Agricultural and Food Chemistry*, vol. 48, no. 10, pp. 4581-9, 2000.
- [34] A. Karaali, N. Şahin, M. Gültekin, "A novel potential ingredient for functional foods: Punicalagins of pomegranates," in *Proc. 2nd International Congress on Functional Foods and Nutraceuticals*, 4-6 May, 2006, Istanbul.
- [35] A. Akpınar-Bayizit, T. Özcan, L. Yılmaz-Ersan, E. Yıldız, and B. Delikanlı, "Functional properties and anti-microbial activity of some macrofungal extracts," in *Proc. International Food Congress-Novel Approaches in Food Industry*, May 26-29, 2011, İzmir, Türkiye, pp. 10-15.
- [36] E. Poyrazoğlu, V. Gökmen, and N. Artık, "Organic acids and phenolic compounds in pomegranates (*Punica granatum* L.) grown in Turkey," *J Food Comp Anal*, vol. 15, pp. 567-575, 2002.
- [37] H. Vardin and M. Abbasoglu, "Nar Eksisi ve Narin Diger Degerlendirme Olanaklari," *Geleneksel Gidalar Sempozyumu*, Van, Türkiye, 2004, 23-24 September.
- [38] A. Kaya and N. Sözer, "Rheological behaviour of sour pomegranate juice concentrates (*Punica granatum* L.)," *International Journal of Food Science and Technology*, vol. 40, no. 2, pp. 223-227, 2005.
- [39] R. P. Singh, K. N. C. Murthy, and G. K. Jayaprakasha, "Studies on the antioxidant activity of pomegranate peel and seed extracts using in vitro models," *J Agric Food Chem*, vol. 50, pp. 81-86, 2002.
- [40] P. S. Negi and G. K. Jayaprakasha, "Antioxidant and antibacterial activities of *Punica granatum* peel extracts," *J Food Sci*, vol. 68, pp. 1473-7, 2003.
- [41] N. P. Seeram, M. Aviram, Y. Zhang, S. M. Henning, L. Feng, M. Dreher, and D. Heber, "Comparison of antioxidant potency of commonly consumed polyphenol-rich beverages in the United States," *J Agric Food Chem*, vol. 56, no. 4, pp. 1415-22, 2008.
- [42] L. H. Zhang, L. L. Li, Y. X. Li, and Y. H. Zhang, "In vitro antioxidant activities of fruits and leaves of pomegranate," *Acta Hort*, vol. 765, pp. 31-34, 2008.
- [43] M. Cam, Y. Hisil, and G. Durmaz, "Classification of eight pomegranate juices based on antioxidant capacity measured by four methods," *Food Chem*, vol. 112, pp. 721-726, 2009.
- [44] G. Mousavinejad, Z. Emam-Djomeh, K. Rezaei, and M. H. H. Khodaparast, "Identification and quantification of phenolic compounds and their effects on antioxidant activity in pomegranate juices of eight Iranian cultivars," *Food Chemistry*, vol. 115, no. 4, pp. 1274-8, 2009.
- [45] Y. Yılmaz, İ. Çelik, and F. Işık, "Mineral composition and total phenolic content of pomegranate molasses," *Journal of Food, Agriculture & Environment*, vol. 5, no. 3&4, pp. 102-104, July-October 2007.
- [46] B. İncedayı, C. E. Tamer, and Ö. U. Çopur, "A research on the composition of pomegranate molasses," *U. Ü. Ziraat Fakültesi Dergisi*, vol. 24, no. 2, pp. 37-47, 2010.
- [47] B. Cemeröğlu, "Meyve ve sebze işleme endüstrisinde temel analiz metodlari," *Biltav Yayınları*, p. 380, Ankara, 1992.
- [48] AOAC 932.12, *Solids (Soluble) in Fruits and Fruit Products. Official Methods of Analysis of AOAC International*, 14<sup>th</sup> edition. Gaithersburg Maryland: AOAC International, 1984.
- [49] V. Uylaşer and F. Başoğlu, *Gıda Analizleri I-II Uygulama Kılavuzu*, Bursa, 2000, pp. 8-13.
- [50] A. Akpınar-Bayizit and E. Yıldız, "The chemical characteristics and antioxidant potential of strawberry tree fruit (*Arbutus unedo* L.)," *Bothalia: A Journal of Botanical Research*, vol. 44, p. 12, 2014.



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